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조선대학교 대학원

의학과

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이 논문을 의학 석사학위신청 논문으로 제출함

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노해정의 석사학위논문을 인준함

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노 해 정

(지도교수: 임 동 윤)

조선대학교 대학원 의학과

부신수질의 AT_1 및 AT_2 수용체 모두가 부신수질의 카테콜아민(CA) 합성 및 티로신 수산화효소의 전사조절을 유지하고 촉진시키는 것으로 알려져 있다(Armando 등, 2004). 한편 AT_2 수용체 작동제인 T_2-(Ang II 4-8)$_2$ 는 사람 부신크롬친화세포의 일차배양세포에서 노르에피네프린, 에피네프린 및 NPY 유리에 대해 아무런 영향을 미치지 못한다고 하였다(Cavadas 등, 2003). 이와같이 부신에서 AT_1 차단제의 CA 분비에 대한 작용에 관해서 일부 논란이 있는 것 같다. 따라서 본 연구의 목적은 흰쥐 부신의 적출관류모델에서 선택성 AT_1 수용체 차단제인 losartan 이 카테콜아민 유리에 영향을 미치는지를 검색하고자 본 연구를 시행하여 다음과 같은 결과를 얻었다. Losartan (5-50 μM)을 부신정맥 내로 90 분간 관류 시 비교적 용량 및 시간 의존적으로 ACh (5.32×10^{-3} M), 고칼륨 (5.6×10^{-2} M), DMPP (10^{-4} M) 및 McN-A-343 (10^{-4} M)에 의한 CA 분비반응을 유의하게 억제하였다. Losartan 자체는 기초 CA 분비량에 영향을 미치지

않았다. 또한, 90 분 동안 15 μM losartan 존재 하에서, L 형 칼슘통로 활성화제인 Bay-K-8644 (10^{-5} M), 세포질의 내형질세망막에서 Ca^{2+}-ATPase 억제제인 cyclopiazonic acid (10^{-5} M), 선택성 나트륨통로 활성화제인 veratridine (10^{-4} M) 및 angiotensin II (Ang II, 100nM)에 의한 CA 분비반응이 뚜렷이 억제되었다. 그러나 고농도의 losartan (150 및 300 μM) 존재 하에서 ACh 의 CA 분비반응은 오히려 대조치에 비해 현저하게 증강되었다.

이와 같은 연구결과를 종합하여 보면, 흰쥐 적출 관류 부신수질에서 비교적 낮은 농도의 losartan 은 콜린성(니코틴 및 무스카린 수용체) 흥분작용 및 막탈분극에 의한 CA 분비작용을 유의하게 억제하였으나. 고농도의 losartan 은 ACh 에 의한 CA 분비반응을 오히려 현저하게 증강시켰다. Losartan 은 흰쥐 부신수질에서 농도에 따라서 콜린 수용체의 작동제와 길항제로서 작용하는 이중작용이 있는 것으로 여겨진다. Losartan 은 작용미상의 CA 분비 증강작용외에, 이러한 losartan 의 CA 분비 억제작용은 흰쥐 적출 부신수질의 크롬친화세포에 있는 AT_1 수용체의 차단작용을 통해 매개되는 것으로 사료되며, 이는 전압의존성 Na^+ 및 Ca^{2+} 이온통로를 통한 세포내로 나트륨 및 칼슘이온의 유입을 차단하고 세포질내 칼슘저장고로부터 칼슘유리를 억제함으로써 나타나는 것으로 생각된다.

I. INTRODUCTION

Generally, angiotensin II (Ang II) is mostly generated from the inactive decapeptide Angiotensin I (Ang I) by angiotensinconverting enzyme (ACE). There are alternative non-ACE pathways to generate Ang II from Ang I or directly from angiotensinogen (Zaman et al., 2002). Because of these alternatives, non-ACE pathways that can generate Ang II, ACE inhibitors may not totally suppress Ang II production. For this reason, Ang II receptor blockers (ARBs) provide more specific and complete blockade of Ang II and prevent its detrimental actions, regardless of how it is generated.

In the adrenal medulla, Ang II releases catecholamines (CA) by a direct action (Livett and Marley, 1993), mediated either by circulating Ang II or by the intrinsic adrenal renin angiotensin system (Livett and Marley, 1993; Plunkett et al., 1985; Phillips et al., 1993). Both AT_1 and AT_2 Ang II receptors are expressed in the adrenal medulla. In the rat, AT_2 receptors predominate, AT_1 receptors representing only 5–10% of the total number of Ang II receptors (Israel et al., 1995). It appears that AT_1 receptor stimulation is most important as a regulatory factor for adrenomedullary CA synthesis and release. First, AT_1 blockade is sufficient to inhibit *in vivo* adrenal CA release by Ang II (Wong et al., 1990). Second, pretreatment with an insurmountable AT_1 antagonist almost completely abolished the hormonal and sympathoadrenal response to the stress of isolation in unfamiliar metabolic cages (Armando et al., 2001). However, isolation stress also produced a substantial increase in adrenomedullary AT_2 receptor binding, which was abolished by pretreatment with the AT_1 receptor antagonist (Armando

et al., 2001). This indicated a possible role of AT_2 receptors in the adrenomedullary response to stress.

The Ang II type 1 receptor (AT_1) antagonist losartan blocked both inhibition and facilitation of secretion by AngII in cultured bovine chromaffin cells (Teschemacher and Seward, 2000), and chronic blockade (losartan) of rennin-angiotensin-aldosterone system (RAS) in rats may decrease the excess sympathetic responses to stress in cardiovascular diseases as well as prevent the likely development of Type II diabetes mellitus (Uresin et al., 2004). In spontaneously hypertensive rats (SHRs), oral administration of AT_1 antagonist (candesartan) can effectively block central actions of Ang II, regulating blood pressure and reaction to stress, and selectively and differentially modulating sympathoadrenal response (Seltzer et al., 2004). Critchley and his colleagues (2004) have found that the angiotensin II type 1 receptor antagonist candesartan, and the angiotensin II converting enzyme inhibitor ramipril, increased basal CA release from the anaesthetized dog's adrenal gland along with decreases in blood pressure. However, it has been demonstrated that AT_2 stimulation induces CA secretion in cultured porcine chromaffin cells (Takekoshi et al., 2001). AT_2 receptors play a role in mediating CA secretion by the adrenal medulla of anesthetized dogs in response to AngII receptor agonist administration *in vivo*. Furthermore, both PD 123319 and CGP 42112 inhibited the increase in adrenal CA secretion induced by local administration of Ang II (Martineau et al., 1999). Worck and his colleagues (1998) have speculated that angiotensin II through binding to both receptor subtypes (both AT_1 and AT_2) facilitates the sympathoadrenal reflex response by actions at several anatomical levels of the

neural pathways involved in the sympathoadrenal reflex response elicited during insulin-induced hypoglycemia in conscious chronically instrumented rats.

Thus, there seems to be some controversy about the effect of AT_1 receptor blockade on the CA secretion from the adrenal gland. The aim of this study therefore was to determine whether losartan, a seletive antagonist of AT_1 receptor, could influence the CA release in the isolated perfused model of the rat adrenal medulla.

II. MATERIALS AND METHODS

Experimental procedure

Male Sprague-Dawley rats, weighing 200 to 300 grams, were anesthetized with thiopental sodium (50 mg/kg) intraperitoneally. The adrenal gland was isolated by the methods described previously (Wakade, 1981). The abdomen was opened by a midline incision, and the left adrenal gland and surrounding area were exposed by the placement of three-hook retractors. The stomach, intestine and portion of the liver were not removed, but pushed over to the right side and covered by saline-soaked gauge pads and urine in bladder was removed in order to obtain enough working space for tying blood vessels and cannulations.

A cannula, used for perfusion of the adrenal gland, was inserted into the distal end of the renal vein after all branches of adrenal vein (if any), vena cava and aorta were ligated. Heparin (400 IU/ml) was injected into vena cava to prevent blood coagulation before ligating vessels and cannulations. A small slit was made into the adrenal cortex just opposite entrance of adrenal vein. Perfusion of the gland was started, making sure that no leakage was present, and the perfusion fluid escaped only from the slit made in adrenal cortex. Then the adrenal gland, along with ligated blood vessels and the cannula, was carefully removed from the animal and placed on a platform of a leucite chamber. The chamber was continuously circulated with water heated at 37 $\pm 1^{o}$C (Fig. 2).

Perfusion of adrenal gland

The adrenal glands were perfused by means of peristaltic pump (ISCO® pump,

WIZ Co. U.S.A.) at a rate of 0.33 ml/min. The perfusion was carried out with Krebs-bicarbonate solution of following composition (mM): NaCl, 118.4; KCl, 4.7; $CaCl_2$, 2.5; $MgCl_2$, 1.18; $NaHCO_3$, 25; KH_2PO_4, 1.2; glucose, 11.7. The solution was constantly bubbled with 95 % O_2 + 5 % CO_2 and the final pH of the solution was maintained at 7.4 ~ 7.5. The solution contained disodium EDTA (10 μg/ml) and ascorbic acid (100 μg/ml) to prevent oxidation of catecholamines.

Drug administration

The perfusions of DMPP (10^{-4} M) and Ang II for 1 or 2 minutes and/or a single injection of ACh (5.32 x 10^{-3} M) and KCl (5.6 x 10^{-2} M) in a volume of 0.05 ml were made into perfusion stream via a three-way stopcock, respectively. McN-A-343 (10^{-4} M), veratridine (10^{-4} M), Bay-K-8644 (10^{-5} M) and cyclopiazonic acid (10^{-5} M) were also perfused for 4 min, respectively.

In the preliminary experiments, it was found that upon administration of the above drugs, the secretory responses to ACh, KCl, McN-A-343, veratridine, Bay-K-8644 and cyclopiazonic acid returned to preinjection level in about 4 min, but the responses to DMPP in 8 min.

Collection of perfusate

As a rule, prior to stimulation with various secretagogues, the perfusate was collected for 4 min to determine the spontaneous secretion of CA (background sample). Immediately after the collection of the background sample, collection of the perfusates was continued in another tube as soon as the perfusion medium containing the stimulatory agent reached the adrenal gland. Stimulated sample's

perfusate was collected for 4 to 8 min. The amounts secreted in the background sample have been subtracted from that secreted from the stimulated sample to obtain the net secretion value of CA, which is shown in all of the figures.

To study the effect of losartan on the spontaneous and evoked secretion, the adrenal gland was perfused with Krebs solution containing losartan for 60 min, and then the perfusate was collected for a certain period (background sample). Then the medium was changed to the one containing the stimulating agent or along with losartan, and the perfusates were collected for the same period as that for the background sample. The adrenal gland's perfusate was collected in chilled tubes.

Measurement of catecholamines

CA content of perfusate was measured directly by the fluorometric method of Anton and Sayre (1962) without the intermediate purification alumina for the reasons described earlier (Wakade, 1981) using fluorospectrophotometer (Kontron Co., Milano, Italy).

A volume of 0.2 ml of the perfusate was used for the reaction. The CA content in the perfusate of stimulated glands by secretagogues used in the present work was high enough to obtain readings several folds greater than the reading of control samples (unstimulated). The sample blanks were also lowest for perfusates of stimulated and non-stimulated samples. The content of CA in the perfusate was expressed in terms of norepinephrine (base) equivalents.

Statistical analysis

The statistical difference between the control and pretreated groups was determined by the Student's *t* and ANOVA tests. A P-value of less than 0.05 was considered to represent statistically significant changes unless specifically noted in the text. Values given in the text refer to means and the standard errors of the mean (SEM). The statistical analysis of the experimental results was made by computer program described by Tallarida and Murray (1987).

Drugs and their sources

The following drugs were used: losartan, cyclopiazonic acid, acetylcholine chloride, 1.1-dimethyl-4-phenyl piperazinium iodide (DMPP), norepinephrine bitartrate, methyl-1,4-dihydro-2, 6-dimethyl-3-nitro-4-(2-trifluoro- methyl-phenyl) -pyridine-5-carboxylate (BAY-K8644), veratridine hydrochloride, angiotensin II (Sigma Chemical Co., U.S.A.), and (3-(m-chloro-phenyl-carbamoyl-oxy)-2-butynyltrimethyl ammonium chloride [McN-A-343] (RBI, U.S.A.). Drugs were dissolved in distilled water (stock) and added to the normal Krebs solution as required except Bay-K-8644, which was dissolved in 99.5 % ethanol and diluted appropriately with Krebs-bicarbonate solution (final concentration of alcohol was less than 0.1 %). Concentrations of all drugs are expressed in terms of molar base.

III. RESULTS

Effects of losartan on CA secretion evoked by ACh, high K^+, DMPP and McN-A-343 from the perfused rat adrenal glands

After the perfusion with oxygenated Krebs-bicarbonate solution for 1 hr, basal CA release from the isolated perfused rat adrenal glands amounted to 22 ± 3 ng for 2 min (n=12). Since a number of previous studies have indicated that the selective blockade of AT_1 receptors failed to abolish the increase in adrenal CA secretion induced by Ang II (Bunn and Marley 1989; Powis and O'Brien 1991; Wong et al., 1990; Martineau et al., 1995), it was attempted initially to examine the effects of losartan itself on CA secretion from the perfused model of the rat adrenal glands. However, in the present study, losartan (10^{-5} ~ 10^{-4} M) itself did not produce any effect on basal CA output from perfused rat adrenal glands (data not shown). Therefore, it was decided to investigate the effects of losartan on cholinergic receptor stimulation- as well as membrane depolarization-mediated CA secretion. Secretagogues were given at 15 min-intervals. Losartan was present for 90 minutes after the establishment of the control release.

When ACh (5.32×10^{-3} M) in a volume of 0.05 ml was injected into the perfusion stream, the amount of CA secreted was 1364±39 ng for 4 min. However, in the presence of losartan in the range of 5 ~ 50 μM for 90 min, ACh-stimulated CA secretion was inhibited in concentration- and time-dependent fashion. As shown in Fig. 2, in the presence of losartan, CA releasing responses were inhibited by 68% of the corresponding control release. Also, the depolarizing agent, high potassium, markedly stimulated the CA secretion

(635±26 ng for 0-4 min). However, following the pretreatment with losartan (5 ~ 50 μM), high K^+ (5.6 x 10^{-2} M)-stimulated CA secretion was significantly inhibited by 63% of the control at last period (90-94 min) as shown in Fig. 3. DMPP (10^{-4} M), which is a selective nicotinic (N_N) receptor agonist in autonomic sympathetic ganglia, evoked a sharp and rapid increase in CA secretion (1317±33 ng for 0-8 min). However, as shown in Fig. 4, DMPP-evoked CA secretion after pretreatment with losartan was greatly reduced to 71% of the control release (100%). McN-A-343 (10^{-4} M), which is a selective muscarinic M_1-receptor agonist (Hammer and Giachetti, 1982), perfused into an adrenal gland for 4 min also caused an increased CA secretion (524±25 ng for 0-4 min). However, in the presence of losartan, McN-A-343-evoked CA secretion was markedly depressed to 67% of the corresponding control secretion (100%) as depicted in Fig. 5.

Effect of losartan on CA secretion evoked by Bay-K-8644, cyclopiazonic acid, veratridine and angiotensin II from the perfused rat adrenal glands

Since Bay-K-8644 is known to be a calcium channel activator, which enhances basal Ca^{2+} uptake (Garcia et al., 1984) and CA release (Lim et al., 1992), it was of interest to determine the effect of losartan on Bay-K-8644-evoked CA secretion from the isolated perfused rat adrenal glands. Bay-K-8644 (10^{-5} M)-evoked CA secretion in the presence of losartan (15 μM) was greatly blocked to 75% of the control at 75-94 min period as compared to the corresponding control release (512±28 ng for 0-4 min) from 7 adrenal glands as shown in Fig. 6.

Cyclopiazonic acid, a mycotoxin from *Aspergillus* and *Penicillium*, has been

described as a highly selective inhibitor of Ca^{2+}-ATPase in skeletal muscle sarcoplasmic reticulum (Goeger and Riley, 1989; Seidler et al., 1989). The inhibitory action of losartan on cyclopiazonic acid-evoked CA secretory response was observed as shown in Fig. 7. In the presence of losartan (15 μM) from 8 adrenal glands, cyclopiazonic acid (10^{-5} M)-evoked CA secretion was also inhibited to 72% of the control response (464±23 ng for 0-4 min).

The voltage-dependent Na^+ channels consist of the principal α-subunit, which is associated with a noncovalently attached β_1-subunits, and a disulfide-linked β_2-subunit (Catterall, 2000). It has also been known that veratridine-induced Na^+ influx mediated through Na^+ channels increased Ca^{2+} influx via activation of voltage-dependent Ca^{2+} channels and produced the exocytotic secretion of CA in cultured bovine adrenal medullary cells (Wada et al., 1985). To characterize the pharmacological action of losartan on voltage-dependent Na^+ channels, the effect of losartan on the CA secretion induced by veratridine was examined here. As shown in Fig. 8, veratridine greatly produced CA secretion (1259±27 ng for 0-4 min). However, in the presence of losartan (15 μM), veratridine (100 μM)-evoked CA secretion was greatly inhibited to 68% of the corresponding control release.

Since Hano and his colleagues (1994) have suggested that Ang II increase epinephrine release from the adrenal medulla via the AT_1 receptors, it was likely interesting to examine the effect of Ang II on the CA rease. Ang II (100 nM) significantly evoked the CA secretory response (469±54 ng for 0-4 min) whereas, in the presence of losartan (15 μM), Ang II (100 nM)-evoked CA secretion was greatly inhibited to 46% of the corresponding control release (Fig. 9).

High dose effects of losartan on CA release evoked by ACh from the perfused rat adrenal glands

As shown in Fig. 2~9, it has also been shown that losartan inhibits the CA secretory response evoked by cholinergic stimulation in the perfused rat adrenal gland. Therefore, in order to study the high dose effects of losartan on the CA secretion, in the presence of high doses (150 and 300 μM) of losartan, the CA secretory responses evoked by ACh-stimulation were examined. In the presence of losartan (150 μM) for 90 min, ACh-evoked CA release was not affected at initial periods (0~49 min), but since then significantly enhanced to 106% of the corresponding control release as illustrated in Fig. 10. Moreover, after treatment with higher concentration (300 μM) for 90 min, ACh-evoked CA release was greatly enhanced to 123% of the corresponding control release during all periods (Fig. 10).

IV. DISCUSSION

These results obtained from the present study suggest that losartan can inhibit the CA secretion evoked by cholinergic stimulation (both nicotininc and muscarinic receptors) and membrane depolarization from the rat adrenal medulla. This inhibitory effect of losartan seems to be mediated by blocking the influx of Na^+ and Ca^{2+} ions through their channels as well as by inhibiting the release of Ca^{2+} from cytoplasmic store through the blockade of Ang II AT1 receptors located on the presynatic membrane of the rat adrenomedullary chromaffin cells, which are relevant to adrenal nicotinic receptor blockade.

In support of the present results, previously immobilisation stress has been shown to cause increase in plasma norepinephrine (NE) and epinephrine (E) levels (Kubo et al., 2001; Saiki et al., 1997). Intracerebroventricular application of ARBs inhibits the increases in plasma NE and E levels during stress exposure, indicating that the central Ang II system has an excitatory role in sympathetic responses to stress (Saiki et al., 1997). Armando and his colleagues (2001) found that pre-treatment with candesartan, an ARB, eliminated the increase in adrenal NE and E concentrations induced by isolation stress. On the other hand, it has been shown that acute and chronic stress stimulates the RAS to increase the levels of Ang II, both in the plasma and brain (Yang et al., 1993). It was also found that isolation stress enhanced Ang II receptor expression to a similar extent as occurs during repeated immobilisation stress (Saavedra, 1992; Aguilera et al., 1995; Castrén et al., 1988). Üresin and his colleagues (2004) have speculated that chronic blockade (losartan) of RAS in rats may decrease the excess

sympathetic responses to stress in cardiovascular diseases and prevent the likely development of Type II diabetes mellitus. The Ang II type 1 receptor (AT_1) antagonist losartan blocked both inhibition and facilitation of secretion by AngII in cultured bovine chromaffin cells. The results of this study show that activation of multiple types of G-proteins and transduction pathways by a single neuromodulator acting through one receptor type can produce concentration-dependent, bi-directional regulation of exocytosis (Teschemacher and Seward, 2000). Based on previous findings, the present results that losartan dose- and time-dependently reduced the CA secretory responses evoked by ACh, high potassium, DMPP and McN-A-343 from the perfused rat adrenal medulla might be due to the blockade of AT_1 receptors located presynaptically on rat adrenomedullary chromaffin cells. Moreover, it has been shown that, in spontaneously hypertensive rats (SHRs), oral administration of AT_1 antagonist (candesartan) can effectively block central actions of Ang II, regulating blood pressure and reaction to stress, and selectively and differentially modulating sympathoadrenal response and the hypothalamic–pituitary–adrenal stimulation produced by brain Ang II—effects of potential therapeutic importance (Seltzer et al., 2004). Barber and his co-workers (1999) have also suggested that, in SHR, AT_2 receptor activation can facilitate the initial depressor response caused by an AT_1 receptor antagonist.

In the present study, as shown in Fig. 9, losartan also greatly inhibited Ang II-evoked CA release from the rat adrenal medulla. This finding indicates that losartan can inhibit the CA release evoked by cholinergic stimulation as well as by membrane depolarization.

On the other hand, it has been demonstrated that, in cultured porcine chromaffin

cells, AT_2 stimulation induces CA secretion by mobilizing Ca^2 through voltage-dependent Ca^2 channels without affecting intracellular pools and that these effects could be mediated by a decrease in cGMP production (Takekoshi et al., 2001). Worck and his colleagues (1998) have also speculated that angiotensin II through binding to both receptor subtypes (both AT_1 and AT_2) facilitates the sympathoadrenal reflex response by actions at several anatomical levels of the neural pathways involved in the sympathoadrenal reflex response elicited during insulin-induced hypoglycemia in conscious chronically instrumented rats.

In contrast, Takekoshi and his co-workers (2001) have demonstrated that CGP 42112 (AT_2-R agonist) reduces both TH-enzyme activity and TH-synthesis biosynthesis in cultured porcine adrenal medullary cells and that these inhibitory effects could be mediated by decrease of cGMP production. Moreover, Martineau and his co-workers (1999) have suggested that AT_2 receptors play a role in mediating CA secretion by the adrenal medulla of anesthetized dogs in response to AngII receptor agonist administration *in vivo*. PD 123319 and CGP 42112 were devoid of any agonist actions with respect to CA output by the adrenal gland *in vivo*. Furthermore, both PD 123319 and CGP 42112 inhibited the increase in adrenal CA secretion induced by local administration of Ang II.

In light of these results, the present findings seem to be disagreement with those results that adrenal CA secretion is mediated through AT_2 receptors.

Armando and his colleagues (2004) have demonstrated that both adrenomedullary AT_1 and AT_2 receptor types maintain and promote the adrenomedullary CA synthesis and the transcriptional regulation of TH in rats. Instead of opposing effects, however, these results indicate a complex synergistic

regulation between the AT_1 and AT_2 receptor types.

Jezova and his co-workers (1998) have also supported the hypothesis of an AT_1/AT_2 receptor cross-talk in the rat adrenomedullary ganglion cells, and a role for both receptor types on the selective regulation of basal NE, but not E formation, and on the regulation of basal TH transcription. Whereas AT_1 and AT_2 receptors involve the Fos-related antigen Fra-2, AT_1 receptor transcriptional effects include pCREB and ERK2, indicating common as well as different regulatory mechanisms for each receptor type.

The nicotinic receptor is a neurotransmitter-gated cation-conducting ion channel that is opened by binding of agonists such as ACh and DMPP (McGehee and Role, 1995). The opening of this channel triggers Ca^{2+} uptake and secretion of CA from chromaffin cells (Wada et al., 1985). To determine if the inhibition of DMPP-stimulated secretion by AT_1 antagonist was due to an effect on the activity of the nicotinic receptor, the effect of losartan, an AT_1-selective agonist, on DMPP-stimulated CA secretion was examined. As shown in Fig. 4, treatment with losartan greatly inhibited DMPP-evoked CA secretion, reducing by 71% of the control release. The present data are similar to the result that chronic immobilization stress increased plasma glucose, NE, E and corticosterone levels in the rats, and that the ARB losartan significantly prevented these increments induced by chronic stress when given before the stress regimen (Üresin et al., 2004). The role of stress has been implicated in the development of hyperglycaemia by activating pituitary-adrenocortical and sympathoadrenal systems (Kemmer et al., 1986). It has been reported that psychological stress-related variables and insulin resistance are correlated (Raikkonen et al.,

1996) and decreases in stress exposure are associated with better glycaemic control in Type II diabetes mellitus (Van der Does et al., 1996).

It is likely plausible that losartan can activate a signal transduction pathway that is altering the activity of both nicotinic receptors and voltage-sensitive Na^+ channels. It has been shown that most of AngII's physiological effects, such as those exerted on the cardiovascular system and fluid volume homeostasis, are mediated by AT_1; these effects are linked to 1,4,5-inositol triphosphate (IP_3) production after phospholipase C activation, resulting in mobilization of intracellular Ca^2 (Timmermans et al., 1993). Activation of such a pathway could result in elevated levels of Ca^{2+}, diacylglycerol, and inositol trisphosphate in the cells. Consequently Ca^{2+}-dependent and protein kinase C (PKC)-dependent pathways may be activated. PKC has been reported to attenuate the activity of both nicotinic receptors (Swope et al., 1992) and voltage-sensitive Na^+ channels (Catterall, 1992). Thus, these previous findings are in accordance with the present results that losartan inhibited the CA secretion evoked by ACh, DMPP and veratridine.

In the present study, losartan, an AT_1-selective antagonist inhibited the CA secretory responses by high potassium, a direct membrane depolarizer, as well as by Bay-K-8644, an activator of L-type Ca^{2+} channels, which facilitates the influx of Ca^{2+} into the cells. The observation that AT_1-selective antagonist inhibited the CA secretion evoked by Bay-K-8644 was surprising, as Takekoshi et al. (2001) have reported that removal of external Ca^2 significantly suppressed either AngII plus CV-11974 (AT_1 antagonist, 100 nM; which simulates specific AT_2 stimulation) or CGP 42112 (AT_2 agonist)-induced CA secretion in cultured porcine adrenomedullary chromaffin cells. It is unclear how the blockade of AT_1 receptors

results in the inhibition of secretion seen in these cells. The simplest interpretation is that the decrease in Ca^{2+} uptake by losartan is responsible for the observed inhibition of the CA secretion. However, such an interpretation is complicated by the complexity of the relationship between the CA secretion and intracellular free Ca^{2+} levels. Both the intracellular location of the Ca^{2+} level increase (Cheek, 1989; Ghosh and Greenberg, 1995) and the magnitude of the Ca^{2+} level increase (Holz et al., 1982) can affect the relationship between intracellular free Ca^{2+} levels and secretion. Holz et al. (1982) have reported that when Ca^{2+} uptake is large, changes in Ca^{2+} uptake resulted in less than proportional changes in CA secretion. Consequently, although the decrease in Ca^{2+} uptake (influx) into the adrenal chromaffin cells may explain the decrease by losartan in CA secretion, it is still unclear whether this is only or even most important factor contributing to the inhibition of CA secretion by the AT_1 antagonist. However, in view of the results so far obtained from the present study, it is felt that the voltage-sensitive Ca^{2+} channels located on chromaffin cell membrane of the rat adrenal medulla could be the target site for losartan-mediated inhibition of CA secretion.

In the present study, losartan also inhibited the CA secretory responses evoked by cyclopiazonic acid, which is known to be a highly selective inhibitor of Ca^{2+}-ATPase in skeletal muscle sarcoplasmic reticulum (Goeger & Riley, 1989; Seidler et al., 1989). Therefore, it is felt that the inhibitory effect of losartan on the CA secretion evoked by cholinergic stimulation as well as by membrane-depolarization may be associated with the mobilization of intracellular Ca^{2+} in the chromaffin cells. This indicates that the blockade of AT_1 receptors causes an inhibitory effect on the release of Ca^{2+} from the intracellular pools

induced by stimulation of muscarinic ACh receptors, which is weakly responsible for the secretion of CA. In the present work, losartan time- and concentration-dependently produced the inhibition of CA secretion evoked by McN-A-343, a selective muscarinic M_1-agonist. This fact suggests new other concept that losartan can modulate the CA secretory process induced by activation of muscarinic M_1-receptors as well as neuronal nicotinic receptors in the rat adrenal medulla. In supporting this finding, it has been shown that cyclopiazonic acid easily penetrates into the cytoplasm through the plasma membrane and reduces Ca^{2+}-ATPase activity in sarcoplasmic/endoplasmic reticulum, resulting in increase in the subsequent Ca^{2+} release from those storage sites and thereby increase of Ca^{2+}-dependent K^+-current (Suzuki et al., 1992). Moreover, in bovine adrenal chromaffin cells, stimulation of muscarinic ACh receptors is also proposed to cause activation of phosphoinositide metabolism, resulting in the formation of inositol 1,4,5-trisphosphate, which induces the mobilization of Ca^{2+} from the intracellular pools (Cheek et al., 1989; Challiss et al., 1991). However, in the present study, it is uncertain whether the inhibitory effect of the losartan on Ca^{2+} movement from intracellular pools is due to their direct effect on the PI response or the indirect effect as a result of AT_1 receptor blockade by losartan. Based on these previous results, this finding of the present work suggests that AT_1 receptor blockade-induced inhibition may be involved in regulating CA secretion evoked by muscarinic M_1-receptor stimulation in the rat adrenal medullary chromaffin cells. Furthermore, Ang II is a secretogogue for CA secretion that is believed to be mediated through IP_3 production by AT_1 (Wong et al., 1990; Dendorfer et al., 1998). Indeed, Wong and his colleagues (1990) demonstrated that AngII-induced CA release is mediated

by AT_1 in the rat adrenal medulla. AT_1-mediated phospholipase C activation and subsequent IP_3 formation may increase cytosolic Ca^2 levels by releasing Ca^2 from intracellular storage, with subsequent activation of CA release (Israel et al., 1995). Indeed, it has been shown that addition of IP_3 to permeabilized bovine chromaffin cells releases intracellular Ca^2 (Stoehr et al., 1986). Furthermore, addition of Ca^2 to permeabilized bovine chromaffin cells was reported to cause CA secretion (Dunn and Holz, 1983).

On the other hand, in the present work, high concentrations of losartan (150 and 300 μM) significantly enhanced ACh-evoked CA secretory responses. As this result alone, there seems to be difficult for interpretation of the enhancement of ACh-evoked CA secretion by high dose of losartan.

In support of this idea, the research results of Vijayapandi and Nagappa (2005) showed biphasic effects of losartan ptassium on immobility in mice: reducd immobility at lower dose (0.1 and 5 mg/kg, i.p.) and enhanced immobility in higher dose (100 mg/kg, i.p.). These biphasic effects were further confirmed by interaction of losartan potassium with reserpine and antidepressant drugs, nortriptylline and fluoxetine (Vijayapandi and Nagappa, 2005). Nahmod and his colleagues (1978) found Ang II to cause 5-HT release and accelerate its synthesis in biphasic manner, stimulating at high doses and inhibiting at lower doses.

Activation of AT_2 receptor seems to induce effects opposite to that of AT_1 (Unger, 1999). AT_2 stimulation inhibits drinking responses and vasopressin release following centrally administered Ang II (Hohle et al., 1995), promotes differentiation and axonal regeneration, and inhibits proliferation of neuronal cells (Gendron et al., 1999). Thus the counteracting effects between AT_1 and AT_2

receptors suggest that a negative cross talk exist between the AT_1 and AT_2 receptors (Horiuchi et al., 1999), as is the case in catecholaminergic neurons (Gelband et al., 1997). Most of the central effects of Ang peptides, which are mediated by AT_1 receptor, are under control by AT_2 receptor. They are in accordance with earlier findings from in vitro experiments in endothelial cells where growth promoting effects mediated by AT_1 receptors were counteracted by growth inhibitory actions of AT_2 receptors (Stoll et al., 1995). Opposite effects of AT_1 and AT_2 receptors on the second messenger phosphatidylinositol have also been described by Gyurko and his co-workers (1992). AT_1 selective receptor antagonists are known to bind to AT_2 receptors with low affinity and vice versa. However, the selectivity is not absolute, and large concentrations of AT_1 selective receptor antagonists can displace Ang II from AT_1 receptors to the alternative site (AT_2 receptors) (Saavedra et al., 1999). Vijayapandi and Nagappa (2005) have obtained that the biphasic effect of losartan potassium on immobility in mice might be due to inhibitory effect on AT_1 receptor at lower dose and pronounced effect on AT_2 receptor at higher dose (large concentrations of losartan potassium can displace Ang II from its AT_1 receptor to AT_2 receptor). In chronic studies with losartan potassium even at lower dose (3 mg/kg, P.O.) potentiated immobility in mice, which might be due to continuous blockade of AT_1 receptor resulting in unopposed AT_2 receptor stimulation (Vijayapandi and Nagappa, 2005). It has also been previously reported that the treatment of Ang II for 4 h has a biphasic effect on Na^+ transport in the primary cultured rabbit renal proximal tubule cells (PTCs) ; a pico molar range of Ang II stimulates Na^+ transport, whereas a micro molar range of Ang II inhibits it (Han et al., 2000). Based on these previous results, in the present study, it seems that biphasic effects of losartan on the CA secretion in

the perfused rat adrenal medulla are due to inhibitory effect on AT_1 receptor at lower dose (5~50 μM) and pronounced effect on AT_2 receptor at higher dose (150 and 300 μM), indicating that large concentrations of losartan can displace Ang II from its AT_1 receptor to AT_2 receptor. However, the detailed relationship between AT_1 and AT_2 receptors in adrenomedullary CA secretion should be confirmed in the future study.

As shown in Fig. 10, taken together, these experimental results suggest that losartan at low concentrations inhibits the CA secretion evoked by cholinergic stimulation (both nicotininc and muscarinic receptors) as well as by membrane depolarization from the rat adrenal medulla, but at high concentration it rather inhibits Ach-evoked CA secretion. It seems that losartan has dual action acting as both agonist and antagonist at nicotinic receptors of the rat adrenal medulla, which might be dependent on the concentration. It is also thought that this inhibitory effect of losartan may be mediated by blocking the influx of both Na^+ and Ca^{2+} through their channels into the rat adrenomedullary chromaffin cells as well as by inhibiting the Ca^{2+} release from its cytoplasmic calcium store, which is thought to be relevant to AT_1 receptor blockade, in addition to its unknown enhancement effect on the CA release.

V. SUMMARY

Armando and his colleagues (2004) demonstrated that both adrenomedullary AT_1 and AT_2 receptor types maintain and promote the synthesis of adrenomedullary catecholamines (CA) and the transcriptional regulation of TH. On the other hand, it has been found that the receptor-AT_2 agonist, T_2-(Ang II 4–8)$_2$ has no effect on NE, EP and NPY release from primary cultures of human adrenal chromaffin cells (Cavadas et al, 2003). Thus, there seems to be some controversy about the effect of AT_1 receptor blockade on the CA secretion from the adrenal gland. The aim of this study therefore was to determine whether losartan could influence the CA release from the isolated perfused model of the rat adrenal medulla. Losartan (5~50 μM) perfused into an adrenal vein for 90 min produced dose- and time-dependently inhibited the CA secretory responses evoked by ACh (5.32 mM), high K^+ (56 mM, a direct membrane depolarizer), DMPP (100 μM) and McN-A-343 (100 μM). Losartan failed to affect basal CA output. Furthermore, in adrenal glands loaded with losartan (15 μM) for 90 min, the CA secretory responses evoked by Bay-K-8644 (10 μM, an activator of L-type Ca^{2+} channels), cyclopiazonic acid (10 μM, an inhibitor of cytoplasmic Ca^{2+}-ATPase), veratridine (100 μM, an activator of Na^+ channels), and angiotensin II (Ang II, 100nM) were markedly inhibited. However, at a high concentration (300 μM), losartan rather enhanced the CA secretion evoked by ACh. Collectively, these experimental results suggest that losartan at low concentrations inhibits the CA secretion evoked by cholinergic stimulation (both nicotininc and muscarinic receptors) as well as by membrane depolarization from

the rat adrenal medulla, but at high concentration it rather inhibits Ach-evoked CA secretion. It seems that losartan has dual action acting as both agonist and antagonist at nicotinic receptors of the rat adrenal medulla, which might be dependent on the concentration. It is also thought that this inhibitory effect of losartan may be mediated by blocking the influx of both Na^+ and Ca^{2+} into the rat adrenomedullary chromaffin cells as well as by inhibiting the Ca^{2+} release from the cytoplasmic calcium store, which is thought to be relevant to AT_1 receptor blockade, in addition to its enhancement effect on the CA release.

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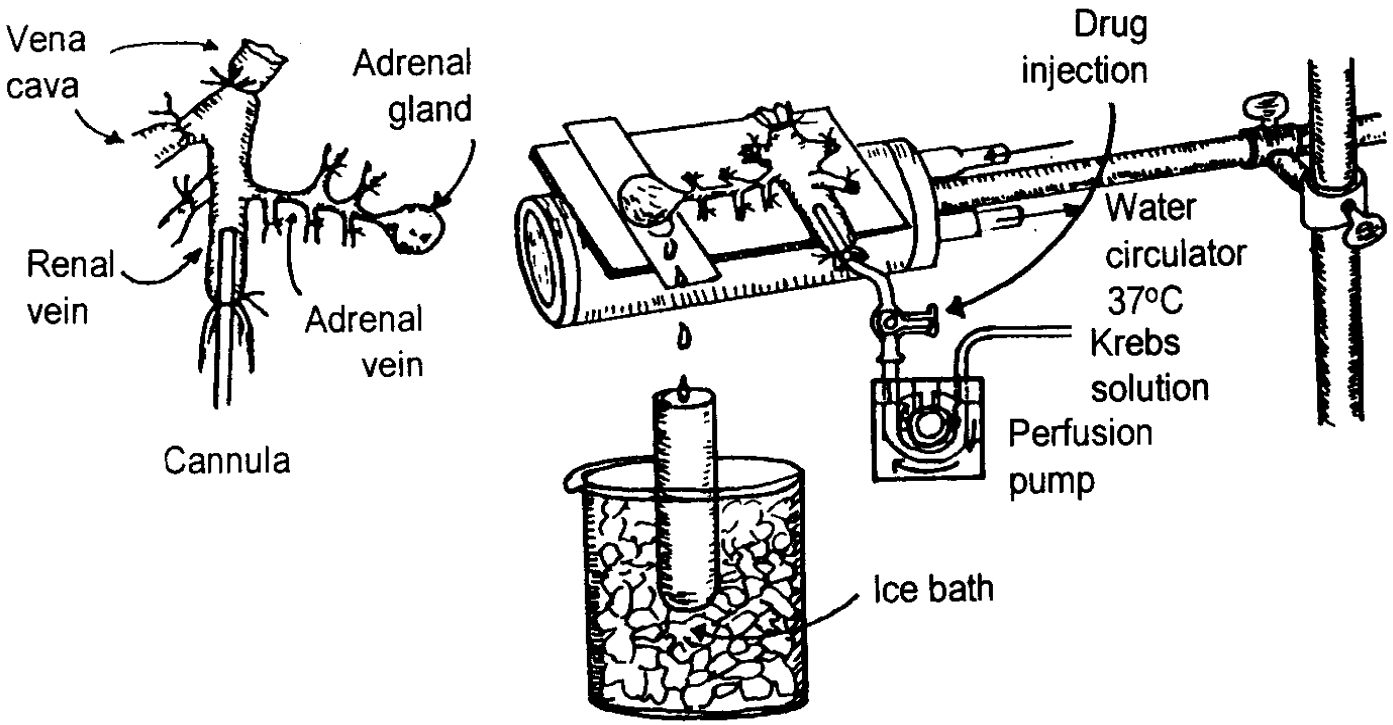

Fig. 1. Schematic drawing of the preparation used to study secretion of catecholamines in the isolated perfused rat adrenal gland.

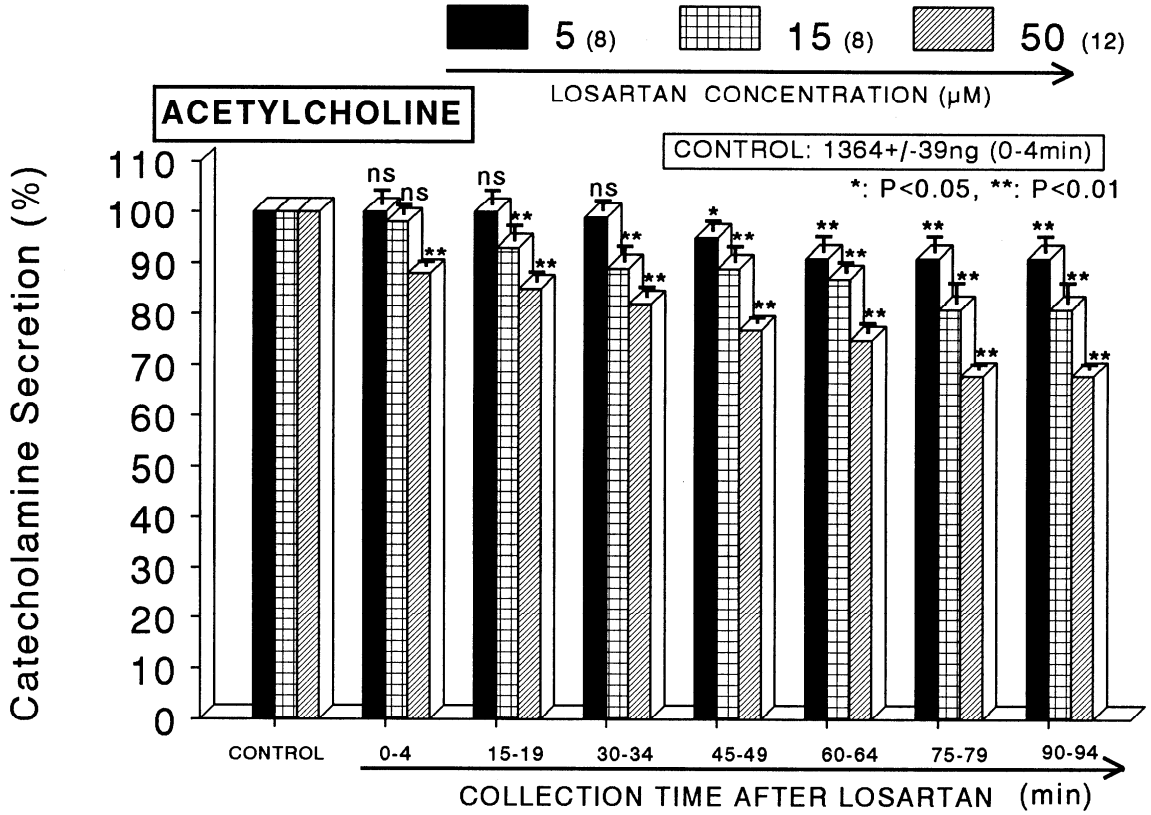

Fig. 2. Dose-dependent effects of losartan on the secretory responses of catecholamines (CA) evoked by acetylcholine (ACh) from the isolated perfused rat adrenal glands. CA secretion by a single injection of ACh (5.32 x 10^{-3} M) in a volume of 0.05 ml was evoked at 15 min intervals after preloading with 5, 15, and 50 μM of losartan for 90 min as indicated at an arrow mark. Numbers in the parenthesis indicate number of rat adrenal glands. Vertical bars on the columns represent the standard error of the mean (S.E.M.). Ordinate: the amounts of CA secreted from the adrenal gland (% of control for 4 min). Abscissa: collection time of perfusate (min). Statistical difference was obtained by comparing the corresponding control (CONTROL) with each concentration-pretreated group of losartan. ACh-induced perfusate was collected for 4 minutes. . *: P < 0.05, **: P < 0.01. ns; Statistically not significant

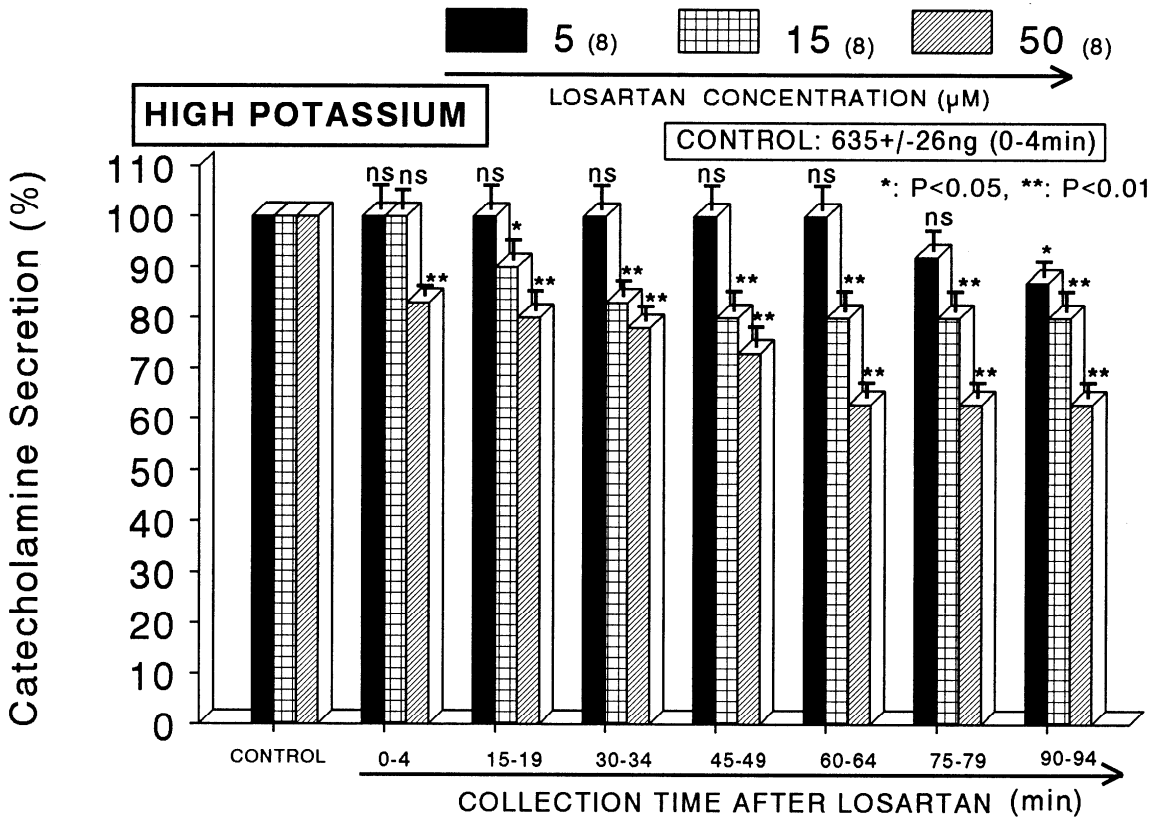

Fig. 3. Dose-dependent effects of losartan on the secretory responses of catecholamines (CA) evoked by high K^+ from the isolated perfused rat adrenal glands. CA secretion by a single injection of K^+ (56 mM) was injected in a volume of 0.1 ml at 15 min intervals after preloading with 5, 15, and 50 μM of losartan for 90 min, respectively. Statistical difference was obtained by comparing the corresponding control (CONT) with each concentration-pretreated group of losartan. K^+-induced perfusate was collected for 4 minutes. Other legends are the same as in Fig. 2. *: $P < 0.05$, **: $P < 0.01$. ns; Statistically not significant.

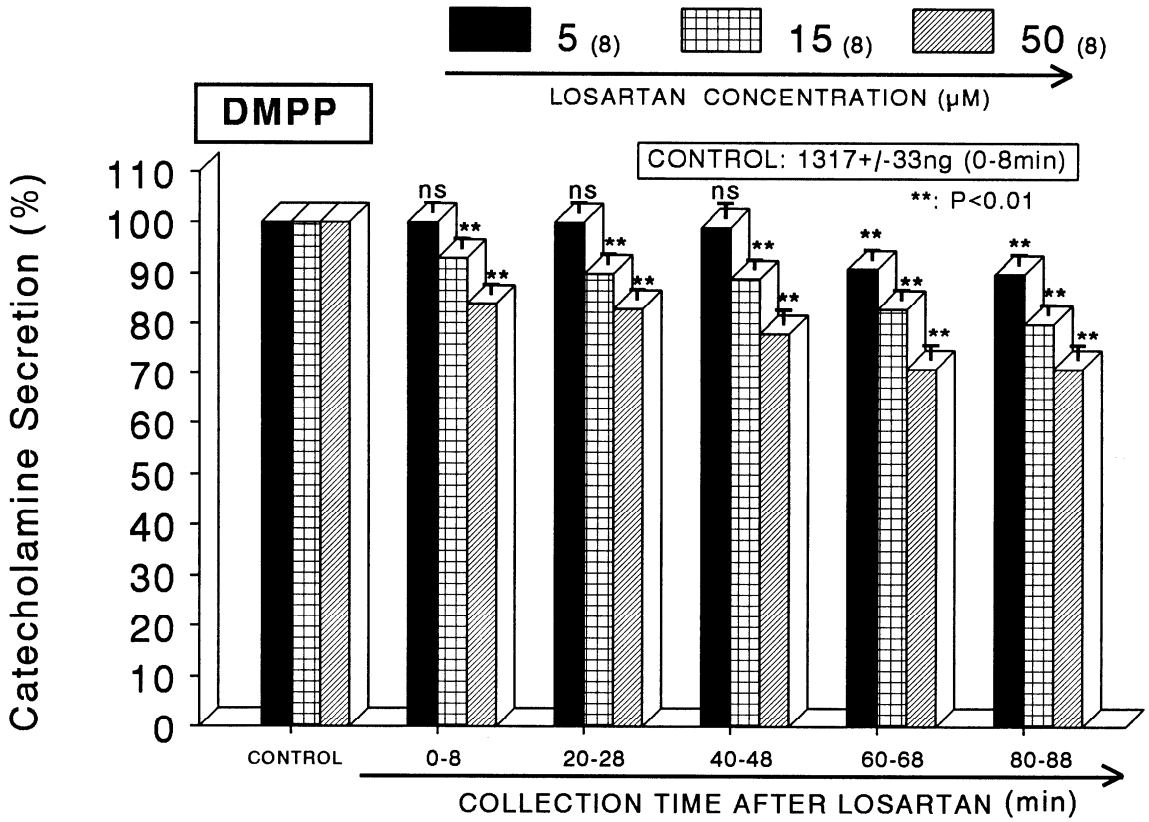

Fig. 4. Dose-dependent effects sof losartan on the secretory responses of catecholamines (CA) evoked by DMPP from the isolated perfused rat adrenal glands. CA secretion by the perfusion of DMPP (10^{-4} M) was infused for 2 min at 20 min intervals after preloading with 5, 15, and 50 μM of losartan for 90 min, respectively. Statistical difference was obtained by comparing the corresponding control with each concentration-pretreated group of losartan. DMPP-induced perfusate was collected for 8 minutes. Other legends are the same as in Fig. 2. **: P < 0.01. ns; Statistically not significant.

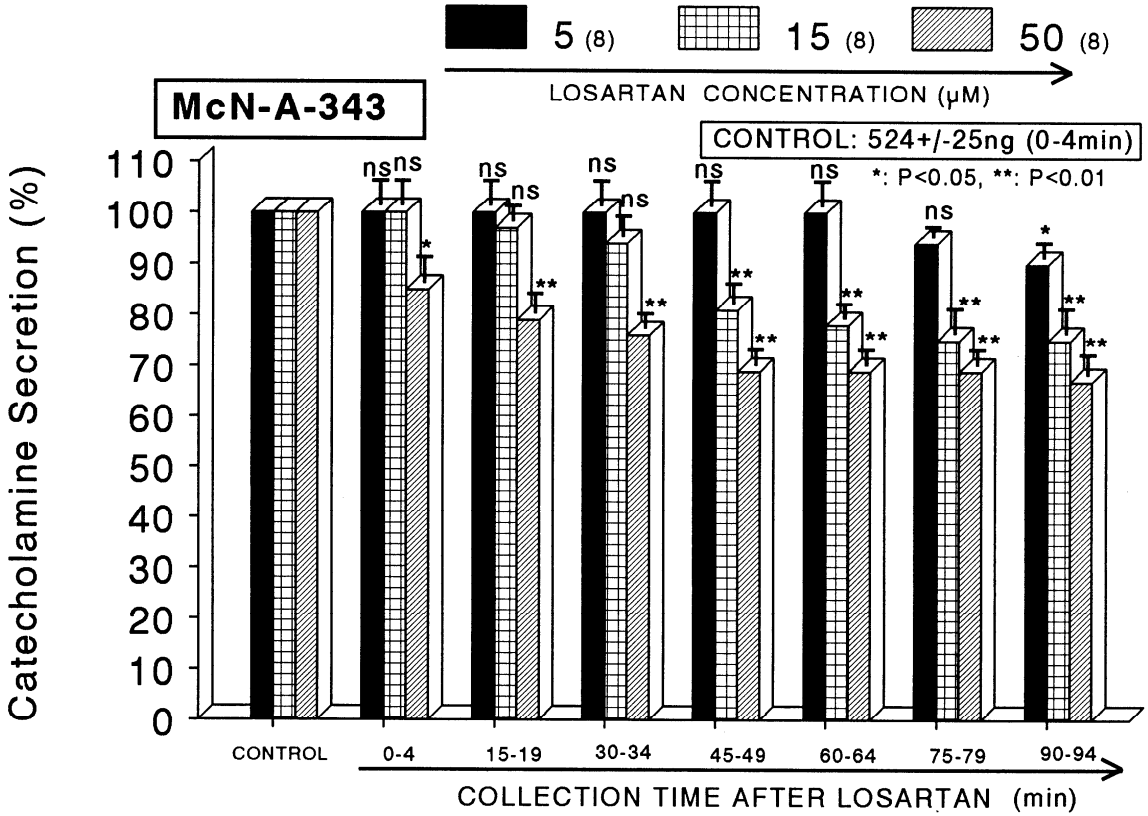

Fig. 5. Dose-dependent effects of losartan on the secretory responses of catecholamines (CA) evoked by McN-A-343 from the isolated perfused rat adrenal glands. CA secretion by the perfusion of McN-A-343 (10^{-4} M) was infused for 4 min at 15 min intervals after preloading with 5, 15, and 50 μM of losartan for 90 min, respectively. Statistical difference was obtained by comparing the corresponding control with each concentration-pretreated group of losartan. McN-A-343-induced perfusate was collected for 4 minutes. Other legends are the same as in Fig. 2. *: $P < 0.05$, **: $P < 0.01$. ns; Statistically not significant.

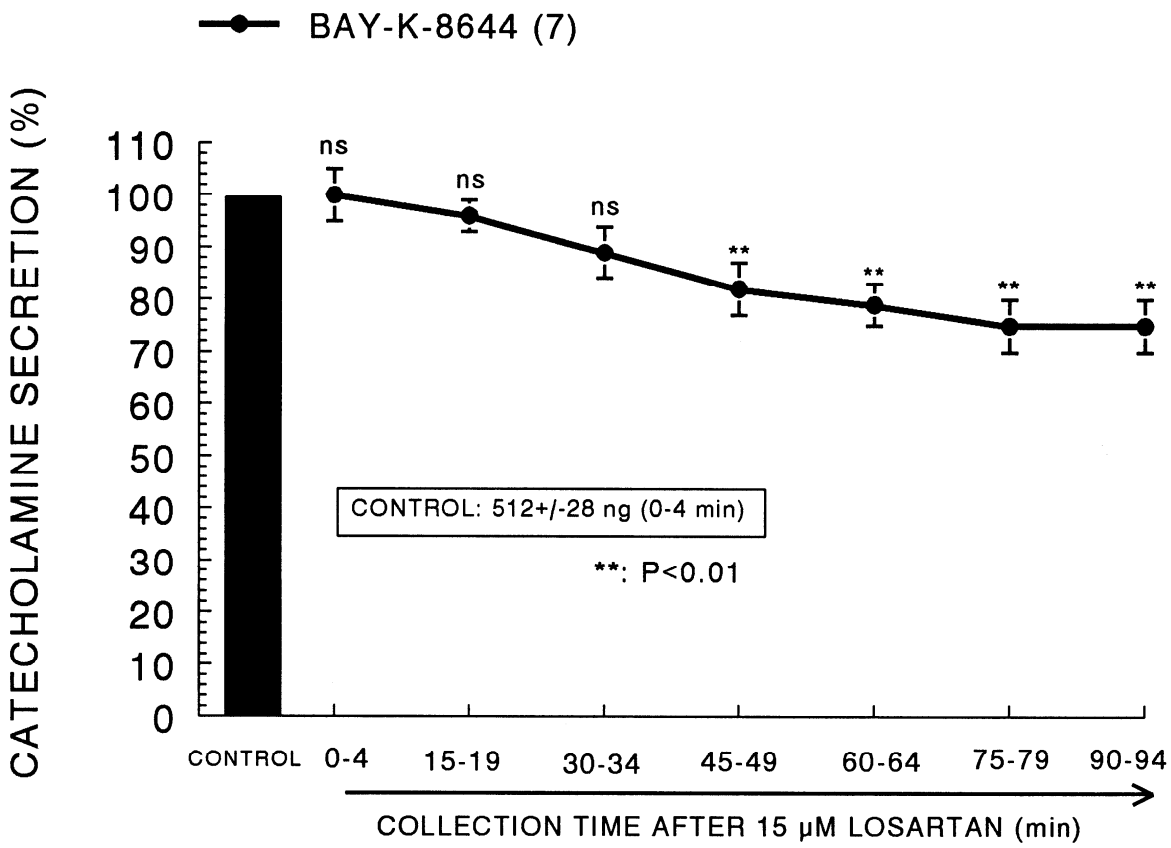

Fig. 6. Time-course effects of losartan on CA release evoked by Bay-K-8644 from the rat adrenal glands. Bay-K-8644 (10^{-5} M) was perfused into an adrenal vein for 4 min at 15 min intervals after preloading with losartan (15 μM) for 90 min, respectively. Statistical difference was obtained by comparing the corresponding control with each period after pretreatment with losartan. Other legends are the same as in Fig. 2. **: P < 0.01. ns; Statistically not significant.

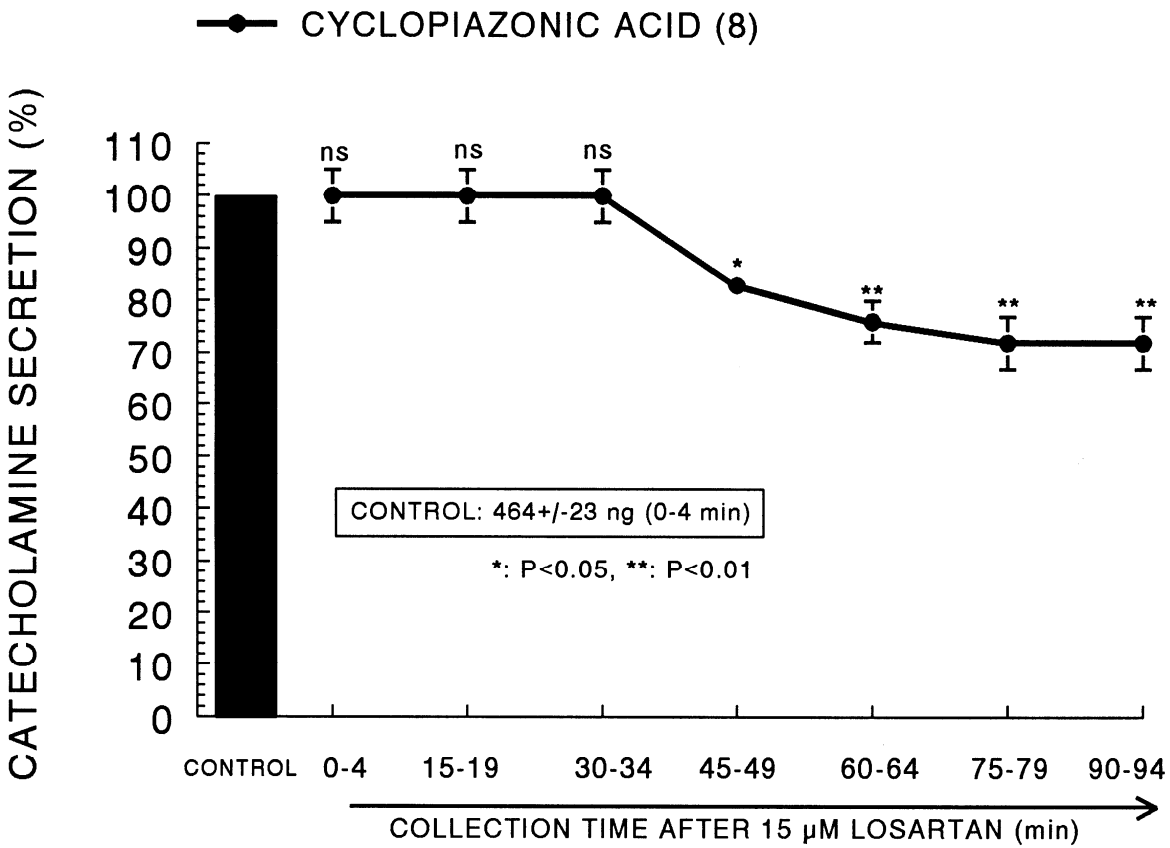

Fig. 7. Time-course effects of losartan on CA release evoked by cyclopiazonic acid from the rat adrenal glands. Cyclopiazonic acid (10^{-5} M) were perfused into an adrenal vein for 4 min at 15 min intervals after preloading with losartan (15 μM) for 90 min, respectively. Statistical difference was obtained by comparing the corresponding control with each period after pretreatment with losartan. Other legends are the same as in Fig. 2. . *: $P < 0.05$, **: $P < 0.01$. ns; Statistically not significant.

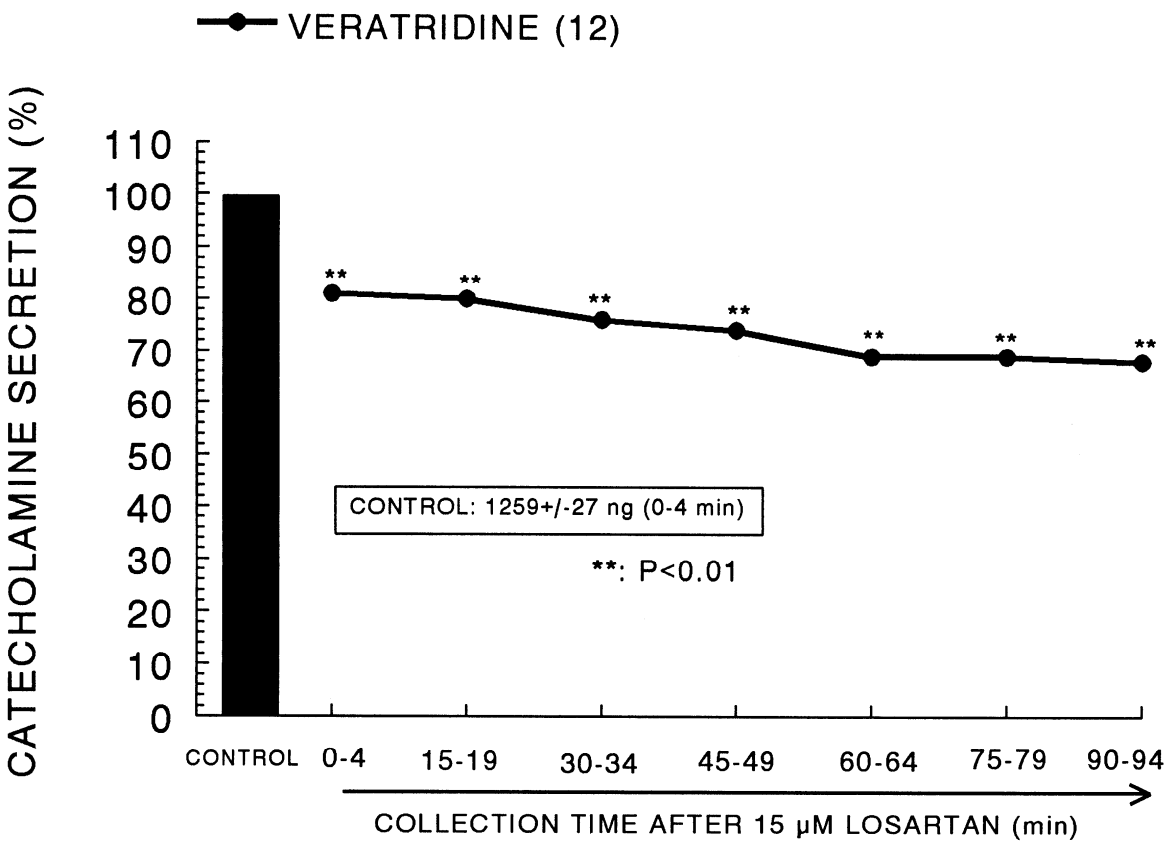

Fig. 8. Time-course effects of losartan on the CA release evoked by veratridine from the rat adrenal glands. Veratridine (10^{-4} M) was perfused into an adrenal vein for 4 min at 15 min intervals after preloading with losartan (15 μM) for 90 min, respectively. Other legends are the same as in Fig. 2. **: P < 0.01.

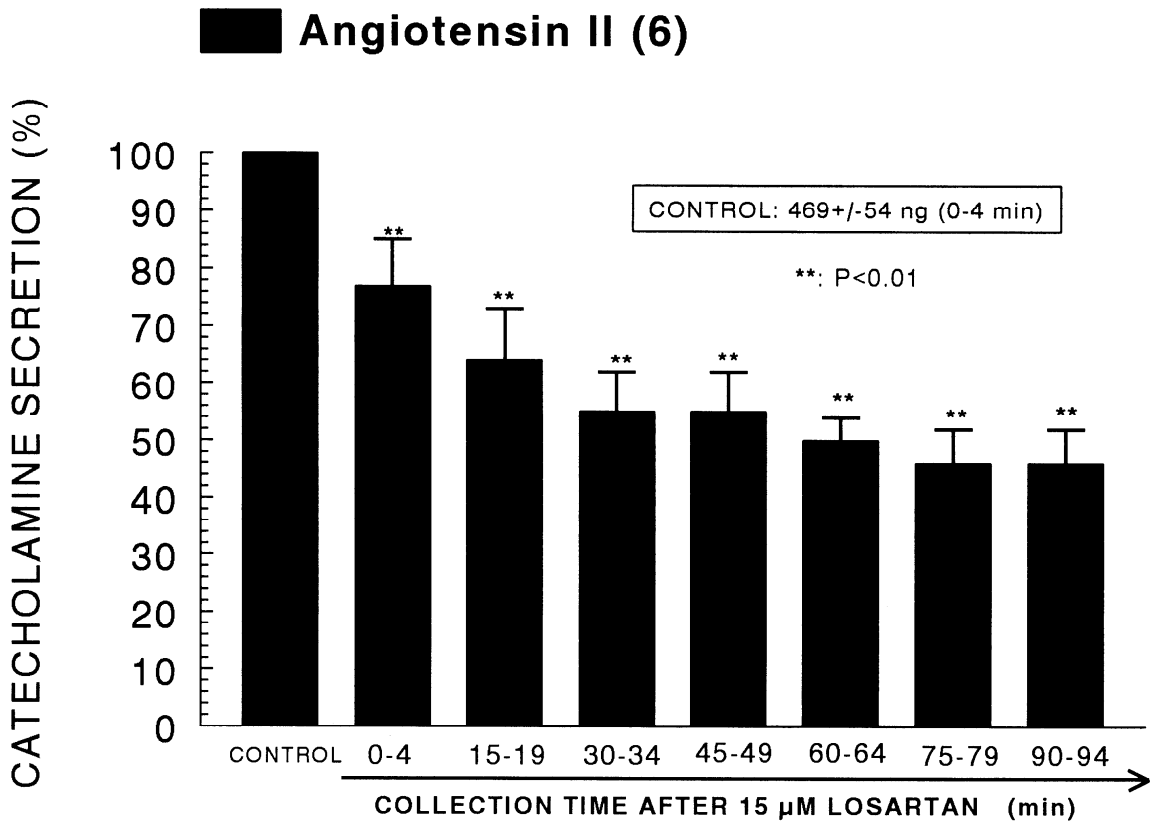

Fig. 9. Time-course effects of losartan on the CA release evoked by angoitensin II from the rat adrenal glands. Angotensin II (10^{-6} M) was perfused into an adrenal vein for 1 min at 15 min intervals after preloading with losartan (15 μM) for 90 min, respectively. Other legends are the same as in Fig. 2. **: P < 0.01.

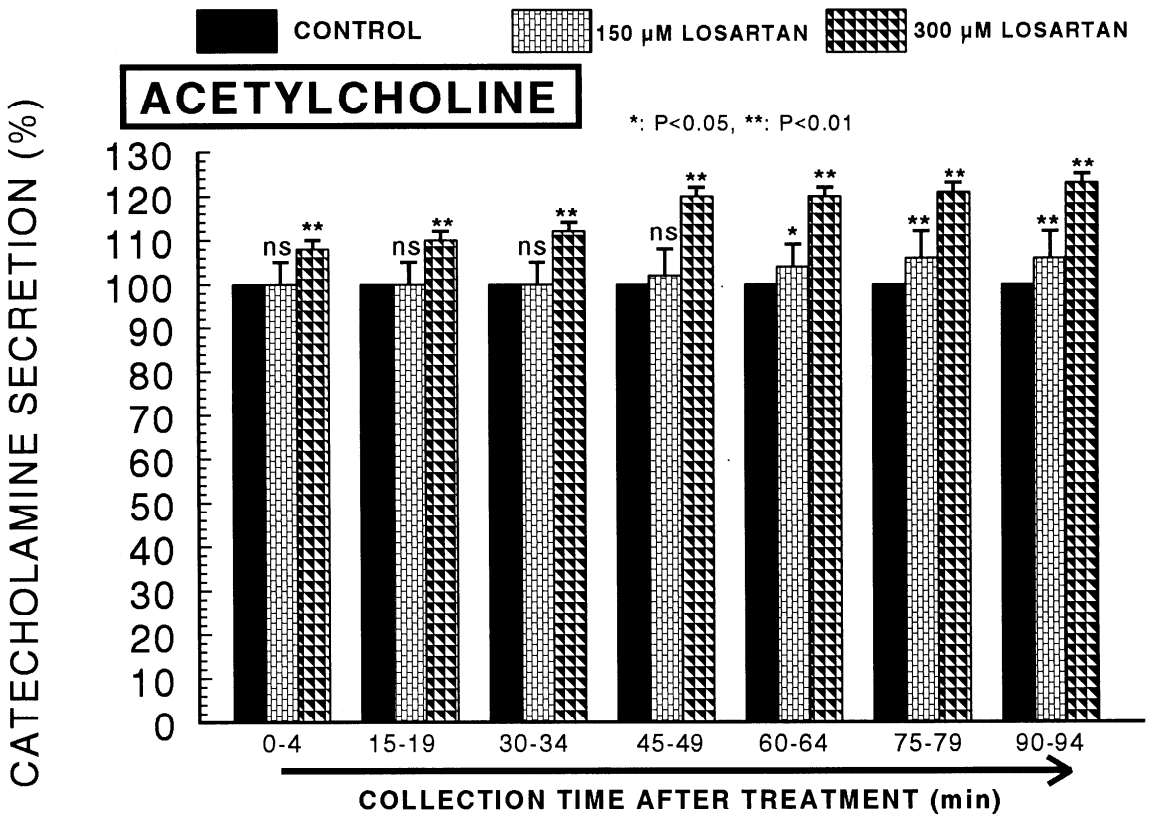

Fig. 10. High dose-effects of losartan on the secretory responses of catecholamines (CA) evoked by acetylcholine (ACh) from the isolated perfused rat adrenal glands. CA secretion by a single injection of ACh (5.32×10^{-3} M) in a volume of 0.05 ml was evoked at 15 min intervals after preloading with 150 and 300 μM of losartan for 90 min as indicated at an arrow mark. ACh-induced perfusate was collected for 4 minutes. Other legends are the same as in Fig. 2. *: $P < 0.05$, **: $P < 0.01$. ns; Statistically not significant.

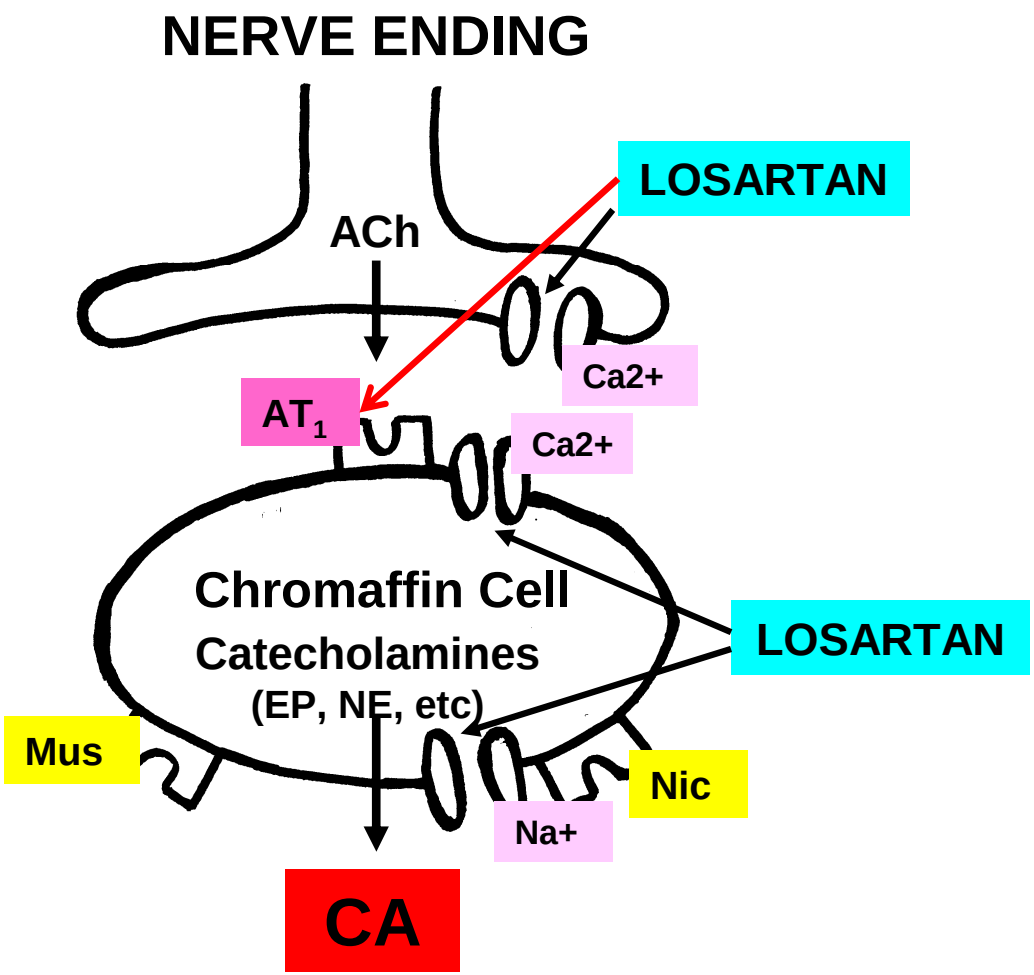

Fig. 10. Schematic diagram of possible action site of losartan at the cholinergic nerve ending-chromaffin cell synapse in the the rat adrenal gland.

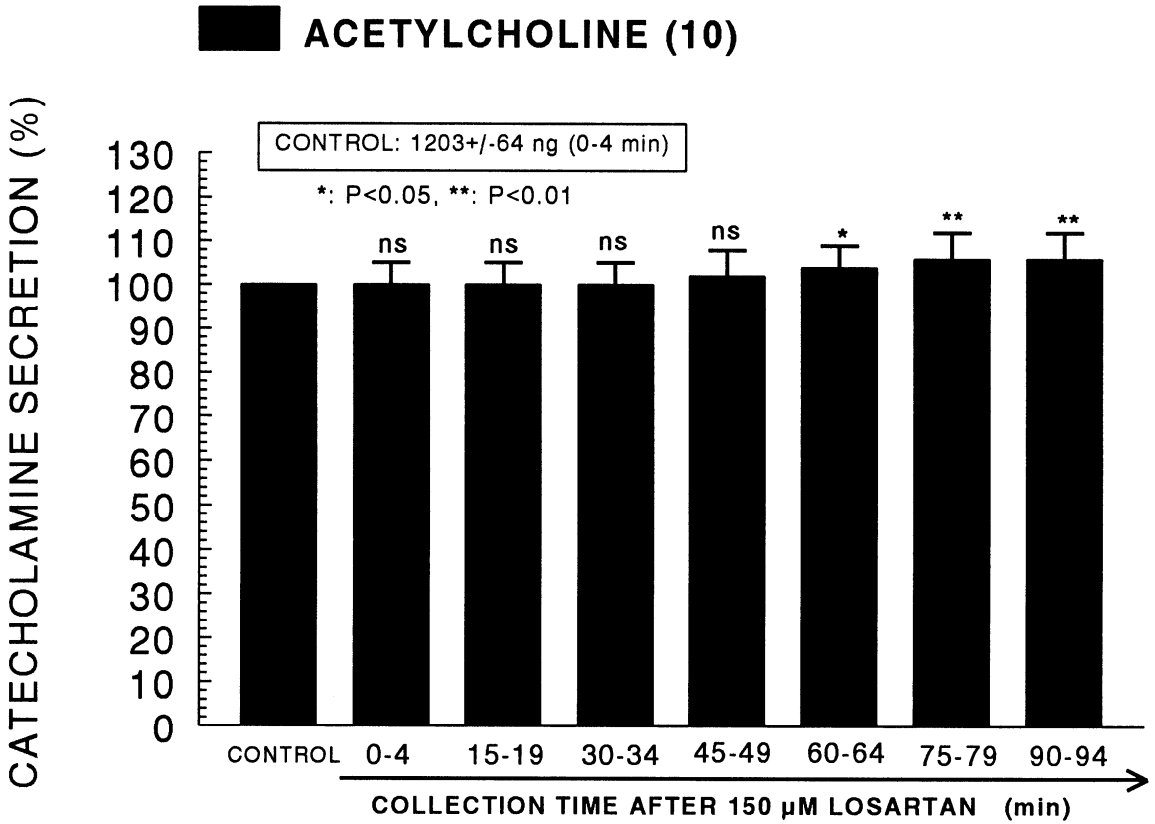

Fig. 10. High dose effect of losartan on the secretory responses of catecholamines (CA) evoked by acetylcholine (ACh) from the isolated perfused rat adrenal glands. CA secretion by a single injection of ACh (5.32×10^{-3} M) in a volume of 0.05 ml was evoked at 15 min intervals after preloading with 150 μM of losartan for 90 min as indicated at an arrow mark. Other legends are the same as in Fig. 3. *: $P < 0.05$, **: $P < 0.01$. ns; Statistically not significant.

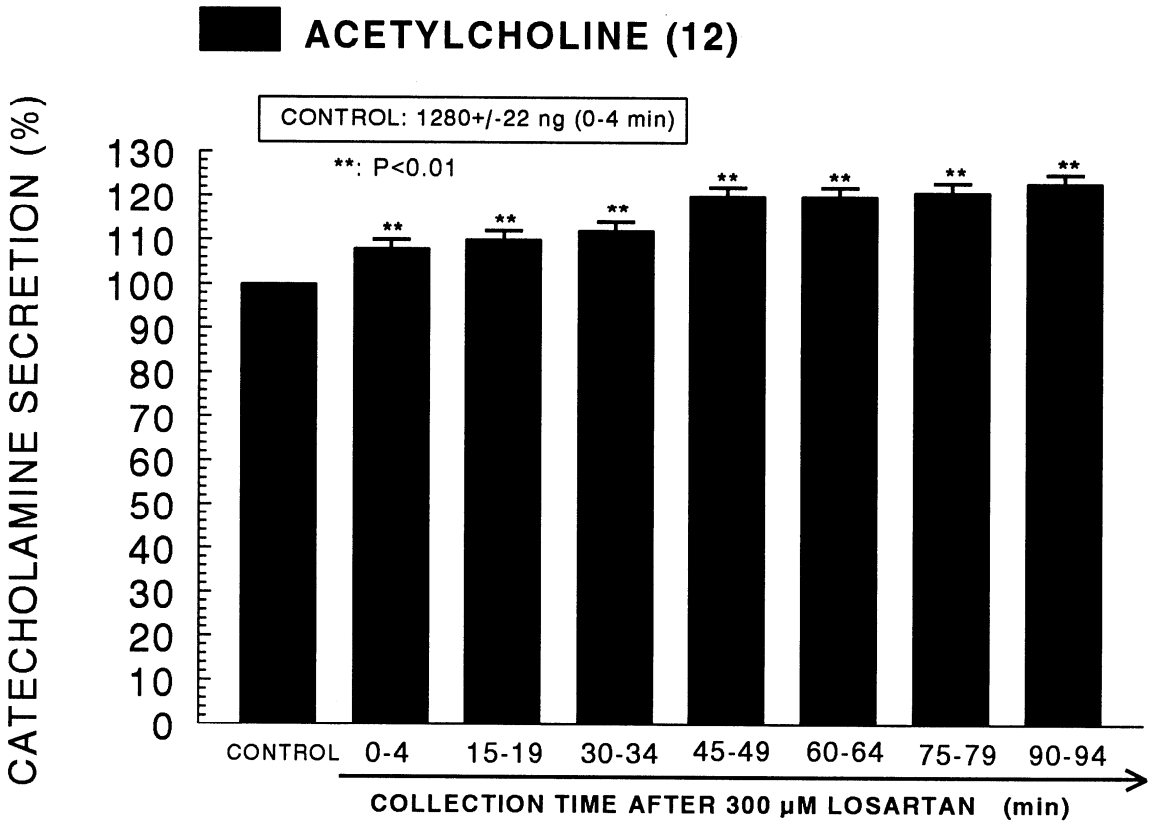

Fig. 11. High dose effect of losartan on the secretory responses of catecholamines (CA) evoked by acetylcholine (ACh) from the isolated perfused rat adrenal glands. CA secretion by a single injection of ACh (5.32×10^{-3} M) in a volume of 0.05 ml was evoked at 15 min intervals after preloading with 300 μM of losartan for 90 min as indicated at an arrow mark. Other legends are the same as in Fig. 3. ACh-induced perfusate was collected for 4 minutes. **: $P < 0.01$.

저작물 이용 허락서

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논문제목	한글: Losartan이 흰쥐 관류 부신수질에서 카테콜아민 분비에 미치는 영향 영어 :Influence of Losartan on Catecholamine Secretion in the Perfused Rat Adrenal Medulla				

본인이 저작한 위의 저작물에 대하여 다음과 같은 조건아래 조선대학교가 저작물을 이용할 수 있도록 허락하고 동의합니다.

- 다 음 -

1. 저작물의 DB구축 및 인터넷을 포함한 정보통신망에의 공개를 위한 저작물의 복제, 기억장치에의 저장, 전송 등을 허락함
2. 위의 목적을 위하여 필요한 범위 내에서의 편집·형식상의 변경을 허락함. 다만, 저작물의 내용변경은 금지함.
3. 배포·전송된 저작물의 영리적 목적을 위한 복제, 저장, 전송 등은 금지함.
4. 저작물에 대한 이용기간은 5년으로 하고, 기간종료 3개월 이내에 별도의 의사 표시가 없을 경우에는 저작물의 이용기간을 계속 연장함.
5. 해당 저작물의 저작권을 타인에게 양도하거나 또는 출판을 허락을 하였을 경우에는 1개월 이내에 대학에 이를 통보함.
6. 조선대학교는 저작물의 이용허락 이후 해당 저작물로 인하여 발생하는 타인에 의한 권리 침해에 대하여 일체의 법적 책임을 지지 않음
7. 소속대학의 협정기관에 저작물의 제공 및 인터넷 등 정보통신망을 이용한 저작물의 전송·출력을 허락함.

동의여부 : 동의(0) 반대( )

2008년 11월 28일

저작자: 노 해 정 (서명 또는 인)

조선대학교 총장 귀하